

INSTRUCTIONS FOR USE Imegen[®] Vegan ID Kit Ref. IMG-230

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Guide overview

The information in this guide is subject to change without notice.

Health in Code, S.L. guarantees that its products are free from defects, both in used materials as in its manufacturing process. This warranty is extended to the expiration date, as long as the storage conditions specified in this manual are met. Our products are designed for research use only. The user of the product is responsible for validating the usefulness of the protocol proposed by Health in Code, S.L. These protocols are considered a guide only. Health in Code, S.L. does not offer any other warranty, express or implied, which extends beyond the proper functioning of the components of this set. Health in Code S.L., sole obligation in respect of the preceding guarantees, will be to replace the product or return the purchase price thereof, as desired by the customer, as long as the existence of a defect in the materials test, or in the manufacture of its products. Health in Code, S.L. will not be responsible for any damage, direct or indirect, resulting in economic losses or damages resulting from the use of this product by the purchaser or user.

All products sold by Health in Code, S.L. are subjected to rigorous quality control (App. A). The **Imegen® Vegan ID Kit** has passed all internal validation tests, ensuring the reliability and reproducibility of each assay.

For any questions about the applications of this product or its protocols, please contact our Technical Department:





tech.support@healthincode.com

NOTE: ImegenAgro® is a trademark of Health in Code, S.L.

Instructions for Use (IFU) modifications

| V.07 | OCT 2024 | Transcription errors: In section 1.3 modification of the positive control's description and table 5 title. Content revision in section 2.1; content added in section 2.3. |
|------|----------|--|
| V.06 | SEP 2023 | Contents review; modification of the storage temperature of the General Master Mix |
| V.05 | AUG 2022 | Change of the manufacturer's identification, going from Imegen to Health in Code, S.L. |
| V.04 | APR 2021 | Contents review |

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Product information

01.1 General description

The identification of meat species presence in food samples is an essential step in order to verify the origin of the used and traceability of the used raw materials, as well as to evaluate the quality control for the handling and cleaning process of production lines by which it passed.

The Health in Code species ID product works by PCR amplification of a specific tag located in the mitochondrial genome of the evaluated species. The particularity of this procedure is due to the fact that the mitochondrial genome is an ideal target since it can be analysed to ensure a specific detection of the desired species and at the same time excluding the detection of other related species. Since there are multiple copies of the mitochondrial genome in each cell, the sensitivity that this detection will have is up to 100 times greater than a test that only target a specific locus in a single copy within the nuclear DNA genome.

During the last decade there had been publicly known cases related to the deceptions that consumers suffer at the time of buying meat and receive other species that are not the ones that they are paying for. EU authorities revealed the presence of uncontrolled meat in food products, and because of this, the food industry authorities have developed food safety management systems to improve the resilience of supply chain to food fraud, mostly directed to prevent the fraud opportunity. Despite the fact, food fraud does not impose a health hazard., but in some ways, they are more dangerous because the raw materials and quality control actions are unknown and untraceable.

The possibility to have a fast and accurate method to determine the authenticity of the ingredient used for food preparation is now available but the precision of the results will be something important to take in consideration at the time of evaluating the food processing. The importance of this phenomenon also lies in economic and commercial problems for both the consumer and the production company Customers want to be sure about the origin of the product they are consuming, also the concern of the contained risk for health. DNA analysis allows a valuable and conscious identification of plants and animal derivatives, by efficiently detecting contaminations or fraud related to inaccurate declaration on the label of the species constituting the food.

01.2 Intended use

Imegen[®] Vegan ID Kit allows determining the presence of animal DNA in any food by real time PCR.

Animal DNA detection is done by real time PCR using two hydrolysis probes. One of them, labelled with FAM[™] dye, specifically detects one mitochondrial DNA sequence common in all animal species. The second probe is labelled with VIC[™] and detects an Internal Positive Control, which is used to rule out inhibitors in the sample and check the correct functioning of the assay.

01.3 Content and storage conditions of the kit

Imegen® Vegan ID Kit contents the necessary reagents to perform 48 reactions:

| Vegan Master Mix | Master Mix with specific oligonucleotides, fluorophore- labelled hydrolysis probes (FAM [™] probe for Animal DNA detection and VIC [™] probe for the Internal Positive Control detection, IPC), synthetic plasmid including the specific IPC sequence and nuclease-free water. |
|---------------------------|--|
| General Master Mix | Master Mix of PCR with nucleotides, MgCl ₂ , DNA polymerase and buffer needed to carry out RT- PCR. |
| Vegan Positive Control | Represents 0.1% of animal genomic DNA. |

Table 1. IMG-230 Imegen® Vegan ID Kit components and description.

| Reagents | Color indicator | Quantity | Conservation |
|-------------------------|-----------------|----------|--|
| Vegan Master Mix* | Black pad | 360 µl | -20 °C |
| General Master Mix* | White pad | 600 µl | -20 °C upon receipt. 2 - 8 °C after initial use. Store protected from light |
| Vegan Positive control* | Orange cap | 120 µl | -20 °C |

(*) See the expiration date on the box and tubes.



01.4 Equipment, reagents and materials required but not supplied

| Equipment | 0 | Real-Time PCR Thermal Cycler with channels for detection of FAM™ (520 nm) and VIC™ (550 nm) |
|-----------|---|--|
| | Ø | Micropipettes (10 μl, 20 μl and 200 μl) |
| | 0 | Tabletop centrifuge with adaptors for 96 well PCR plates and/or 0.2 ml tubes |
| | 0 | Vortex |
| Materials | Ø | Optical 96-well reaction plates or 0.2 ml optical tubes |
| | 9 | Optical adhesive film for 96 well plates or optical caps for 0.2 ml tubes |
| | 0 | Disposable micropipette filter tips (10 μL , 20 μL and 200 μL) |
| | Ø | 1.5 ml sterile tubes |
| | 9 | Powder-free latex gloves |
| Reagents | 0 | Nuclease-free water |





Methods

02.1 Preparation of the amplification reactions

A PCR master mix should be prepared in order to perform the analysis with <u>Imegen®</u> <u>Vegan ID Kit</u>:

The PCR Master Mix contains:

- Vegan Master Mix
- General Master Mix (2X)

The recommended protocol for preparation of amplification reactions is shown below:

- 01. Thaw the Vegan Master Mix, the Positive Control vial and samples.
- 02. Vortex each reagent and keep cold.
- **03**. Add into a 1.5 mL tube (one for each PCR master mix preparation), the following reagents (Table 2). To estimate the amount of necessary reagents, we recommend to make calculations taking into account the number of samples to be simultaneously analyzed, and then considering one more reaction.

Table 2. Reagents and volumes needed to perform PCR reactions.

| Reagents | Amount per reaction |
|--------------------|---------------------|
| Vegan Master Mix | 7.5 μL |
| General Master Mix | 12.5 μL |

- 04. Vortex and spin the 1.5 mL tube and dispense 20 μ l into corresponding wells.
- 05. Add 5 μ l of each DNA sample at 10 ng/ μ l, 5 μ l of Vegan Positive Control and 5 μ l of the Negative Controls* into the appropriate wells.
- **06**. Cover the well plate with optical film or the tubes with optical cover and spin in the centrifuge.



(*) We strongly recommend using an **extraction negative control** for each run of extractions carried out. This control consists in one tube to which no sample is added, and which is summited to the same extraction process as the other samples. Likewise, we recommended using a **PCR negative control** for each PCR run; this tube contains no DNA but all PCR reagents.

02.2 Settings for the Real-Time PCR program

This kit is compatible with the Real-Time PCR platforms 7500 FAST, StepOne Real-Time PCR System (Thermo Fisher Scientific) and QuantStudio[™] 5 Real-Time PCR system.

Table 3. IMG-230 Imegen® Vegan ID Kit probes and specifications.

| Target | Receptor | Quencher |
|------------|----------|----------|
| Animal DNA | FAM™ | MGB |
| IPC | VIC™ | MGB |

The following instructions should be taken into account in order to setup the amplification program:

- Reaction volume: 25 μL.
- Targets: FAMTM and VICTM.
- In case the quencher has to be defined, select MGB for all probes. If the real time PCR system does not take into account the quenchers, select only the receptors (FAM[™] and VIC[™]).
- If the Real-Time PCR system is a 7500 Fast, a StepOne Real-Time PCR system (Thermo Fisher Scientific) or a QuantStudio[™] 5 Real-Time PCR system, select Quantitation - Standard curve as a type of experiment and include ROX[™] as a reference.
- Ramp rate: standard
- Optimal program:

Table 4. Optimal PCR program.

| Fields Step 1 Enzyme activation | | Step 2 PCR | |
|------------------------------------|-----------------|---------------|------------------------------|
| | | 36 c | cycles |
| No. of cycles | 1 initial cycle | Denaturation | Primers binding/extension |
| Temperature | ire 95°C | | 60°C |
| Time | 10 minutes | 15 seconds | 1 minute * |

(*) Fluorescence detection.

02.3 Analysis of results

To analyze the results, it is recommended to establish the Threshold at 0.1 and to keep the default Baseline value to minimize the residual signal in the detection channels.

| Ct settings | Threshold | 0.1 |
|-------------|-----------|------|
| | Baseline | AUTO |

Before analyzing the samples results, it should be checked if obtained results in controls are as expected:

- Positive control: the result must always be positive in all amplification reactions, both in the FAM[™] channel as VIC[™].
- Segative controls: amplification should only be detected in the VIC[™] channel. In this channel an internal positive control (IPC) is detected, which determines the absence of inhibition in the sample.

| IPC | It must be checked that the IPC (VIC ^{m}) is positive in all samples, with a Ct similar to the Positive Control. A negative result in the IPC indicates the presence of inhibitors in the sample. It should be noted that IPC result may be negative in samples where a lot of animal DNA (FAM ^{m}) is detected, because the PCR reagents are exhausted before amplification of the IPC begins. | |
|------------|---|--|
| Animal DNA | Amplification in the FAM™ channel indicates presence of animal DNA in the sample. | |

It is necessary to check if sample Ct is less than the $Ct_{cut-off}$ in order to determine if one reaction of amplification is positive. Any reaction of amplification with Ct upper than $Ct_{cut-off}$ may be considered as negative. The $Ct_{cut-off}$ is equal than the positive control Ct (0.1%) plus 3.32.

Establish the positive cut-off value for the test samples and assign results:

$$Ct_{(cut-off)} = Ct_{(Positive Control)} + 3.32$$

Table 5. Cut-off values. (1) For fresh or minimally processed meats samples, the cut-off value corresponds approximately 0.01% of animal DNA, when DNA concentration is 10ng/uL.

| Sample Ct value | Sample result |
|--------------------------|---------------|
| $Ct > Ct_{(cut-off)}$ | Negative |
| $Ct \leq Ct_{(cut-off)}$ | Positive |



NOTE: Any sample with a Ct equal than Ct_{cut-off} contains approximately 0.01% of animal DNA.

In samples where no amplification in the FAM[™] channel is seen, we can conclude that no animal DNA is detected or that their amount in the sample is below than the detection limit.

The following table shows graphically the results that may be obtained from one sample analysis, as well as the interpretation that should be done from the obtained result:

| Vegan Ma | aster Mix | | |
|--------------------------|-----------------------|--|--|
| Animal (FAM™ channel) | IPC (VIC™ channel) | Interpretation | |
| - | + | No animal DNA is detected | |
| + | + | Animal DNA is detected | |
| - | - | PCR inhibitors presence in the sample* | |
| + | - | Sample with big amount of animal DNA | |

Table 6. Results interpretation.

(*) If presence of inhibitors in the sample is detected, we recommend checking whether there has been an excess of DNA in the reaction (the recommended maximum is 250 ng). If the amount of DNA is right, we recommend repeating DNA extraction. If the problem persists, please contact our technical department.





Troubleshooting

The following table shows graphically the results that may be obtained from the analysis of different assay controls, as well as the interpretation that should be done from the obtained result:

| | Vegan Master Mix | | | |
|--------------------------------|------------------|-----|--|--|
| Controls | Animal | IPC | Interpretation | |
| | + | + | Expected result | |
| Positive control | - | - | ¹ PCR Amplification Failure | |
| Extraction | - | + | Expected result | |
| Extraction Negative Control | + | + | ² Contamination in the DNA extraction procedure | |
| PCR Negative | - | + | Expected result | |
| Control | + | + | ³ PCR contamination with animal DNA | |

(1) PCR Amplification Failure: check amplification program and configuration of fluorescence capture. Amplification failure may be due to a setup technical problem or PCR inhibition.

(2) Contamination in the DNA extraction procedure: contamination may be due to some error made in the process of sample handling, reagents contamination, or environmental contamination. Check DNA extraction protocol, wipe the laboratory where DNA extraction process was performed and take care to avoid any contamination during sample homogenization. If necessary, use new aliquots of the reagents used in DNA extraction.

(3) PCR contaminations with Animal DNA: contamination of PCR reactions may be due to an error made in the process of sample handling, contamination of the reagents or environmental contamination. Thoroughly clean the laboratory where the PCR process was performed, as well as equipment. If necessary, use new aliquots of the reagents used in the PCR. Prepare the PCR reaction containing the Positive Control last to avoid cross contamination.





Limitations

04.1 Equipment

Imegen® Vegan ID Kit has been validated using the following Real-Time PCR systems:

- 7500 FAST Real-Time PCR System (Thermo Fisher Scientific)
- StepOne[™] Plus Real-Time PCR System (Thermo Fisher Scientific)
- QuantStudio5[™] Real-Time PCR System (Thermo Fisher Scientific)

Technically, this kit is compatible with any Real-Time PCR systems that enable the detection of the fluorescence emitted by FAM[™] and VIC[™] fluorophores.

If a PCR system different from the systems described in this section is going to be used, it is possible that the PCR program might need to be readjusted. In this case, please contact our Technical Support Team for more details.

04.2 Reagents

Imegen® Vegan ID Kit has been validated using the reagents included in the kit and the DNA polymerase recommended by the supplier of the Real-Time PCR systems used in the validation as follows:

TaqMan Environmental Master Mix 2.0 (Thermo Fisher Scientific)

If a PCR master mix (DNA polymerase) different from the DNA polymerase used in the validation is going to be used to perform the analysis, a validation with the new reagents is recommended beforehand. Please, contact our Technical Support Team if you request any further information.



04.3 Product Stability

The optimal analytical functioning of this product is confirmed as long as the recommended storage conditions are applied as specified on Section 01.3 (Contents and Storage Conditions) from the reception of the kit until the expiry date assigned to each production batch.



Supplemental information

AA.1 Sensitivity and specificity

The specificity of the kit was tested through comparison with the NCBI sequence database and was also experimentally tested with success on a collection of reference DNAs. See the results in the table below:

Table 8. Specificity of IMG-230 Imegen® Vegan ID Kit.

| Meat species | Result |
|---|----------|
| Horse <i>(Equus caballus)</i> | Detected |
| Mule <i>(Equus asinus x Equus caballus)</i> | Detected |
| Donkey <i>(Equus asinus)</i> | Detected |
| Beef <i>(Bos taurus)</i> | Detected |
| Pork <i>(Sus scrofa domestica)</i> | Detected |
| Water buffalo <i>(Bubalus bubalis)</i> | Detected |
| Fallow deer <i>(Dama dama)</i> | Detected |
| Chicken <i>(Gallus gallus)</i> | Detected |
| Turkey <i>(Meleagris gallopavo)</i> | Detected |
| Goat <i>(Capra aegagrus hircus)</i> | Detected |
| Duck (Genus <i>Anas</i>) | Detected |
| Ostrich <i>(Struthio camelus)</i> | Detected |
| Goose <i>(Anser anser)</i> | Detected |
| Sheep <i>(Ovis aries)</i> | Detected |
| Human <i>(Homo sapiens)</i> | Detected |
| Rabbit <i>(Oryctolagus cuniculus)</i> | Detected |
| Dog <i>(Canis familiaris)</i> | Detected |
| Longin tuna <i>(Thunnus alalunga)</i> | Detected |
| European hake (Mertuccius mertuccius) | Detected |

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| European pilchard <i>(Sardina pilchardus)</i> | Detected |
|---|--------------|
| Atlantic salmon <i>(Salmo salar)</i> | Detected |
| Haddock <i>(Melanogrammus aeglefinus)</i> | Detected |
| Pollack <i>(Pollachius pollachius)</i> | Detected |
| Atlantic cod <i>(Gadus morhua)</i> | Detected |
| Mealworms <i>(Tenebrio molitor)</i> | Not detected |
| Buffalo worms <i>(Alphitobius diaperinus)</i> | Not detected |
| Cricket <i>(Acheta domesticus)</i> | Not detected |
| Migratory locust <i>(Locusta migratoria)</i> | Not detected |
| Mussel <i>(Mytilus galloprovincialis)</i> | Not detected |
| Squid <i>(Loligo vulgaris)</i> | Not detected |
| Prawn <i>(Aristaeomorpha foliacea)</i> | Not detected |
| Wheat <i>(Triticum aestivum)</i> | Not detected |
| Soya (Glycine max) | Not detected |
| Maize <i>(Zea mays)</i> | Not detected |
| Tomato <i>(Solanum lycopersicum)</i> | Not detected |
| Rice <i>(Oryza sativa)</i> | Not detected |
| E. coli <i>(Escherichia coli)</i> | Not detected |

AA.2 Detection limit

The detection limit was calculated with standard samples consisting of mixtures of raw animal meat and other species. The <u>Imegen® Vegan ID Kit</u> can detect blends with as little as 0.01% (w/w) of animal meat. The limit of detection in processed samples varies depending on the composition and food processing.

To ensure the representativeness of the results, we recommend the use of a DNA extraction method that allows you to process a large amount of sample (10-20 g). If you do not have a procedure with these features, we recommend the use of Imegen[®] Food Extraction Kit (Ref No.: IMG-262).

AA.3 Quality certifications

- Health in Code, S.L. is certified against the standard UNE-EN ISO 9001:2015 "Quality management systems" for the design, development, manufacture, and commercialization of kits for genetic analysis.
- Health in Code, S.L. is certified against the standard UNE-EN ISO 14001:2015 "Environmental Management Systems" for the design, development, manufacture, and commercialization of kits for genetic analysis.



Safety warnings and precautions

| (!) | Strictly follow the instructions of this manual, especially regarding the handling and storage conditions. |
|---------------|--|
| Ø | Do not pipette by mouth. |
| \oslash | Do not smoke, drink, or eat in areas where specimens or kit reagents are being handled. |
| ⊘ ?0 ?2 | You must properly protect any skin condition, as well as cuts, abrasions and other skin lesions. |
| کې | Send down the drain only those materials found on the safe list. Compounds not listed are not suitable for drain disposal. Use waste containers according to the local legislation and manage their treatment through an authorised waste manager. |
| è | In case of an accidental release of any of the reagents, avoid contact with skin, eyes and mucous membranes and clean with abundant water. |
| + | The materials safety data sheets of all hazardous components contained in this kit are available on request to Health in Code, S.L. |
| & | This product could require the handling of samples and materials of human and animal origin. You should consider all human and animal source materials as potentially infectious and handled in accordance with OSHA Biosafety Level 2 of bloodborne pathogens or must use other relevant biosafety practices for materials containing or suspect that they may contain infectious agents. |
| \bigcirc | Reagents included in this kit are non-toxic, neither explosive, infectious, radioactive, magnetic, corrosive nor environmental polluters. |

| ß | This kit has been validated with specific equipment under certain conditions, which could be different in other laboratories. It is recommended that each laboratory performs an internal validation when the kit is used for the first time. |
|-----|--|
| (!) | The manufacturer is not responsible for the malfunction of the assay when one or more reagents included in the kit are replaced by other reagents not supplied by Health in Code, S.L. |
| ズ | The manufacturer does not guarantee the reproducibility of the assay when the user employs reagents not validated by Health in Code, S.L., considering them equivalent to those provided in the kit. |

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Documentation and support

AC.1 Food safety support

Please, visit our website for the latest services, orders and support information:



Health in Code certificates of analysis and other product documentation:



portal.imegen.es/en/certificate-of-analysis/

AC.2 Customer and technical support

For any questions about the applications of this product or this protocol, please contact our Technical Department:





tech.support@healthincode.com

NOTE: For SDSs for reagents and chemicals from other manufacturers, please contact the appropriate manufacturer.