



INSTRUCTIONS FOR USE

Imegen[®] Ostrich ID Kit

Ref. IMG-244

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Guide overview

The information in this guide is subject to change without notice.

Health in Code, S.L. guarantees that its products are free from defects, both in used materials as in its manufacturing process. This warranty is extended to the expiration date, as long as the storage conditions specified in this manual are met. Our products are designed for research use only. The user of the product is responsible for validating the usefulness of the protocol proposed by Health in Code, S.L. These protocols are considered a guide only. Health in Code, S.L. does not offer any other warranty, express or implied, which extends beyond the proper functioning of the components of this set. Health in Code S.L., sole obligation in respect of the preceding guarantees, will be to replace the product or return the purchase price thereof, as desired by the customer, as long as the existence of a defect in the materials test, or in the manufacture of its products. Health in Code, S.L. will not be responsible for any damage, direct or indirect, resulting in economic losses or damages resulting from the use of this product by the purchaser or user.

All products sold by Health in Code, S.L. are subjected to rigorous quality control (App. A). The **Imegen® Ostrich ID Kit** has passed all internal validation tests, ensuring the reliability and reproducibility of each assay.

For any questions about the applications of this product or its protocols, please contact our Technical Department:



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NOTE: ImegenAgro® is a trademark of Health in Code, S.L.

Instructions for Use (IFU) modifications

| | | |
|-------------|----------|--|
| V.04 | OCT 2023 | Contents review. Modification of the storage temperature of the General Master Mix |
| V.03 | AUG 2022 | Change of the manufacturer's identification, going from Imegen to Health in Code, S.L. |
| V.02 | JUL 2019 | Contents review |

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01

Product information

01.1 General description

The identification of meat species presence in food samples is an essential step in order to verify the origin of the used and traceability of the used raw materials, as well as to evaluate the quality control for the handling and cleaning process of production lines by which it passed.

The Health in Code species ID product works by PCR amplification of a specific tag located in the mitochondrial genome of the evaluated species. The particularity of this procedure is due to the fact that the mitochondrial genome is an ideal target since it can be analyzed to ensure a specific detection of the desired species and at the same time excluding the detection of other related species. Since there are multiple copies of the mitochondrial genome in each cell, the sensitivity that this detection will have is up to 100 times greater than a test that only target a specific locus in a single copy within the nuclear DNA genome.

During the last decade there had been publicly known cases related to the deceptions that consumers suffer at the time of buying meat and receive other species that are not the ones that they are paying for. EU authorities revealed the presence of uncontrolled meat in food products, and because of this, the food industry authorities have developed food safety management systems to improve the resilience of supply chain to food fraud, mostly directed to prevent the fraud opportunity. Despite the fact, food fraud does not impose a health hazard, but in some ways, they are more dangerous because the raw materials and quality control actions are unknown and untraceable.

The possibility to have a fast and accurate method to determine the authenticity of the ingredient used for food preparation is now available but the precision of the results will be something important to take in consideration at the time of evaluating the food processing. The importance of this phenomenon also lies in economic and commercial problems for both the consumer and the production company. Customers want to be sure about the origin of the product they are consuming, also the concern of the contained risk for health. DNA analysis allows a valuable and conscious identification of plants and animal derivatives, by efficiently detecting contaminations or fraud related to inaccurate declaration on the label of the species constituting the food.

01.2 Intended use

[Imegen® Ostrich ID Kit](#) allows determining the presence of DNA of *Struthio camelus* in any food.

Ostrich DNA detection is done by real time PCR using hydrolysis probes. One of them, labelled with FAM™ dye, specifically detects one mitochondrial DNA sequence of *Struthio camelus*. The second probe is labelled with VIC™ and detects an Internal Positive Control, which is used to rule out inhibitors in the sample and check the correct functioning of the assay.

01.3 Content and storage conditions of the kit

[Imegen® Ostrich ID Kit](#) contents the necessary reagents to perform 48 reactions:

| | |
|---------------------------|--|
| Ostrich Master Mix | Master Mix with specific oligonucleotides, fluorophore-labelled hydrolysis probes (FAM™ probe for Ostrich detection and VIC™ probe for the Internal Positive Control detection, IPC), synthetic plasmid including the specific IPC sequence and nuclease-free water. |
| General Master Mix | Master Mix of PCR with nucleotides, MgCl ₂ , DNA polymerase and buffer needed to carry out real-time PCR. |
| Positive Control | DNA sample containing 0.1% of <i>Struthio camelus</i> specific-DNA. |

Table 1. IMG-244 Imegen® Ostrich ID Kit components and description.

| Reagents | Color indicator | Quantity | Conservation |
|---------------------|-----------------|----------|---|
| Ostrich Master Mix* | Black pad | 360 µl | -20 °C |
| General Master Mix* | White pad | 600 µl | -20 °C upon receipt. 2 - 8 °C after initial use. Store protected from light |
| Positive control* | Black cap | 60 µl | -20 °C |

(*) See the expiration date on the box and tubes.

01.4 Equipment, reagents and materials required but not supplied

Equipment

- ✓ Real-Time PCR Thermal Cycler with channels for detection of FAM™ (520 nm) and VIC™ (550 nm)
- ✓ Micropipettes (10 µL, 20 µL and 200 µL)
- ✓ Tabletop centrifuge with adaptors for 96 well PCR plates and/or 0.2 ml tubes
- ✓ Vortex

Materials

- ✓ Optical 96-well reaction plates or 0.2 ml optical tubes
- ✓ Optical adhesive film for 96 well plates or optical caps for 0.2 ml tubes
- ✓ Disposable micropipette filter tips (10 µL, 20 µL and 200 µL)
- ✓ 1.5 ml sterile tubes
- ✓ Powder-free latex gloves

Reagents

- ✓ Nuclease-free water

02

Methods

02.1 Preparation of the amplification reactions

Imegen® Ostrich ID Kit is designed to determine, in a single PCR reaction, the presence or absence of Ostrich DNA and the internal positive control.

The PCR Master Mix contains:

- ➔ **Ostrich Master Mix**
- ➔ **General Master Mix (2X)**

The recommended protocol for preparation of amplification reactions is showed below:

- 01.** Thaw the Beef Master Mix, the Positive Control vial, and samples DNA.
- 02.** Vortex each reagent and keep cold.
- 03.** Add into a 1.5 mL tube (one for each PCR master mix preparation), the following reagents (Table 2). To estimate the amount of necessary reagents, we recommend make calculations taking into account the number of samples to be simultaneously analyzed, and then considering one more reaction, or increase a 10% the volume of each reagent.

Table 2. Reagents and volumes needed to perform PCR reactions.

| Reagents | Amount per reaction |
|--------------------|---------------------|
| Ostrich Master Mix | 7.5 µL |
| General Master Mix | 12.5 µL |

- 04.** Vortex and spin the 1.5 mL tube and dispense 20 µl per well or tube of 0.2 ml.
- 05.** Add 5 µl of each DNA sample at 10 ng/µl, 5 µl of Chicken Positive Control and 5 µl of the Negative Controls* into the appropriate wells.
- 06.** Cover the well plate with optical film or the tubes with optical cover and spin in the centrifuge.

(*) We strongly recommend using an **extraction negative control** for each run of extractions carried out. This control consists in one tube to which no sample is added, and which is submitted to the same extraction process as the other samples. Likewise, we recommended using a **PCR negative control** for each PCR run; this tube contains no DNA but all PCR reagents.

02.2 Settings for the Real-Time PCR program

This kit is compatible with the Real-Time PCR platforms 7500 FAST, StepOne Real-Time PCR System (Thermo Fisher Scientific) and QuantStudio™ 5 Real-Time PCR system.

Table 3. IMG-244 Imegen® Ostrich ID Kit probes and specifications.

| Target | Receptor | Quencher |
|-----------|----------|----------|
| Ostrich-P | FAM™ | MGB |
| IPC | VIC™ | MGB |

The following instructions should be taken into account in order to setup the amplification program:

- + **Reaction volume:** 25 µL.
- + **Targets:** FAM™ and VIC™.
- + In case the quencher has to be defined, select MGB for all probes. If the real time PCR system does not take into account the quenchers, select only the receptors (FAM™ and VIC™).
- + If the Real-Time PCR system is a 7500 Fast, a StepOne Real-Time PCR system (Thermo Fisher Scientific) or a QuantStudio™ 5 Real-Time PCR system, select Quantitation - Standard curve as a type of experiment and include ROX™ as a reference.
- + **Ramp rate:** standard
- + **Optimal program:**

Table 4. Optimal PCR program.

| Fields | Step 1 | Step 2 | |
|---------------|-------------------|--------------|---------------------------|
| | Enzyme activation | PCR | |
| No. of cycles | 1 initial cycle | 36 cycles | |
| | | Denaturation | Primers binding/extension |
| Temperature | 95°C | 95°C | 60°C |
| Time | 10 minutes | 15 seconds | 1 minute * |

(*) Fluorescence detection.

02.3 Analysis of results

To analyze the results, it is recommended to establish the Threshold at 0.1 and to keep the default Baseline value to minimize the residual signal in the detection channels.

| | | |
|--------------------|-----------|------|
| Ct settings | Threshold | 0.1 |
| | Baseline | AUTO |

Before analyzing the samples results, it should be checked if obtained results in controls are as expected:

- **Positive control:** the result must always be positive in all amplification reactions, both in the FAM™ channel as VIC™.
- **Negative controls:** amplification should only be detected in the VIC™ channel. In this channel an internal positive control (IPC) is detected, which determines the absence of inhibition in the sample.

| | |
|------------|--|
| IPC | It must be checked that the IPC (VIC™) is positive in all samples, with a Ct similar to the Positive Control. A negative result in the IPC indicates the presence of inhibitors in the sample. It should be noted that IPC result may be negative in samples where a lot of Ostrich DNA (FAM™) is detected, because the PCR reagents are exhausted before amplification of the IPC begins. |
|------------|--|

| | |
|----------------|---|
| Ostrich | Amplification in the FAM™ channel indicates presence of Ostrich DNA in the sample |
|----------------|---|

It is necessary to check if sample Ct is less than the $Ct_{cut-off}$ in order to determine if one reaction of amplification is positive. Any reaction of amplification with Ct upper than $Ct_{cut-off}$ may be considered as negative. The $Ct_{cut-off}$ is equal than the positive control Ct (0.1%) plus 3.32.

Establish the positive cut-off value for the test samples and assign results:

$$Ct_{(cut-off)} = Ct_{(Positive\ Control)} + 3.32$$

Table 5. Cut-off values. (1) For fresh or minimally processed meats samples, the cut-off value corresponds approximately 0.01% of Ostrich DNA, when DNA concentration is 10ng/uL.

| Sample Ct value | Sample result |
|--------------------------|----------------------|
| $Ct > Ct_{(cut-off)}$ | Negative |
| $Ct \leq Ct_{(cut-off)}$ | Positive |

NOTE: Any sample with a Ct equal than $Ct_{cut-off}$ contains approximately 0.01% of Ostrich DNA.

In samples where no amplification in the FAM™ channel is seen, we can conclude that no Ostrich DNA is detected or that their amount in the sample is below than the detection limit.

The following table shows graphically the results that may be obtained from one sample analysis, as well as the interpretation that should be done from the obtained result:

Table 6. Results interpretation.

| Ostrich Master Mix | | Interpretation |
|------------------------|--------------------|--|
| Ostrich (FAM™ channel) | IPC (VIC™ channel) | |
| - | + | No Ostrich DNA is detected |
| + | + | Ostrich DNA is detected |
| - | - | PCR inhibitors presence in the sample* |
| + | - | Sample with big amount of Beef DNA |

(*) If presence of inhibitors in the sample is detected, we recommend checking whether there has been an excess of DNA in the reaction (the recommended maximum is 250 ng). If the amount of DNA is right, we recommend repeating DNA extraction. If the problem persists, please contact our technical department.

03

Troubleshooting

The following table shows graphically the results that may be obtained from the analysis of different assay controls, as well as the interpretation that should be done from the obtained result:

Table 7. Possible results and their interpretation.

| Controls | Ostrich Master Mix | | Interpretation |
|-----------------------------|--------------------|-----|--|
| | Ostrich | IPC | |
| Positive control | + | + | Expected result |
| | - | - | ¹ PCR Amplification Failure |
| Extraction Negative Control | - | + | Expected result |
| | + | + | ² Contamination in the DNA extraction procedure |
| PCR Negative Control | - | + | Expected result |
| | + | + | ³ PCR contamination with Ostrich DNA |

(1) PCR Amplification Failure: check amplification program and configuration of fluorescence capture. Amplification failure may be due to a setup technical problem.

(2) Contamination in the DNA extraction procedure: contamination may be due to some error made in the process of sample handling, reagents contamination, or environmental contamination. Check DNA extraction protocol, wipe the laboratory where DNA extraction process was performed and take care to avoid any contamination during sample homogenization. If necessary, use new aliquots of the reagents used in DNA extraction.

(3) PCR contaminations with Ostrich DNA: contamination of PCR reactions may be due to an error made in the process of sample handling, contamination of the reagents or environmental contamination. Thoroughly clean the laboratory where the PCR process was performed, as well as equipment. If necessary, use new aliquots of the reagents used in the PCR. Prepare the PCR reaction containing the Positive Control last to avoid cross contamination.

04

Limitations

04.1 Equipment

[Imegen® Ostrich ID Kit](#) has been validated using the following Real-Time PCR systems:

- 7500 FAST Real-Time PCR System (Thermo Fisher Scientific)
- StepOne™ Plus Real-Time PCR System (Thermo Fisher Scientific)
- QuantStudio™ 5 Real-Time PCR system (Thermo Fisher Scientific)

Technically, this kit is compatible with any Real-Time PCR systems that enable the detection of the fluorescence emitted by FAM™ and VIC™ fluorophores.

If a PCR system different from the systems described in this section is going to be used, it is possible that the PCR program might need to be readjusted. In this case, please contact our Technical Support Team for more details.

04.2 Reagents

[Imegen® Ostrich ID Kit](#) has been validated using the reagents included in the kit and the DNA polymerase recommended by the supplier of the Real-Time PCR systems used in the validation as follows:

- TaqMan Environmental Master Mix 2.0 (Thermo Fisher Scientific)

If a PCR master mix (DNA polymerase) different from the DNA polymerase used in the validation is going to be used to perform the analysis, a validation with the new reagents is recommended beforehand. Please, contact our Technical Support Team if you request any further information.

04.3 Product Stability

The optimal analytical functioning of this product is confirmed as long as the recommended storage conditions are applied as specified on Section 01.3 (Contents and Storage Conditions) from the reception of the kit until the expiry date assigned to each production batch.



Supplemental information

AA.1 Sensitivity and specificity

The specificity of the kit was tested through comparison with the NCBI sequence database and was also experimentally tested with success on a collection of reference DNAs. See the results in the table below:

Table 8. Specificity of IMG-244 Imegen® Ostrich ID Kit.

| Meat species | Result |
|--|--------------|
| Sheep (<i>Ovis aries</i>) | Not detected |
| Goat (<i>Capra aegagrus hircus</i>) | Not detected |
| Horse (<i>Equus caballus</i>) | Not detected |
| Beef (<i>Bos taurus</i>) | Not detected |
| Pork (<i>Sus scrofa domestica</i>) | Not detected |
| Water buffalo (<i>Bubalus bubalis</i>) | Not detected |
| Fallow deer (<i>Dama dama</i>) | Not detected |
| Chicken (<i>Gallus gallus</i>) | Not detected |
| Turkey (<i>Meleagris gallopavo</i>) | Not detected |
| Duck (<i>Genus Anas</i>) | Not detected |
| Ostrich (<i>Struthio camelus</i>) | Detected |
| Goose (<i>Anser anser</i>) | Not detected |
| Human (<i>Homo sapiens</i>) | Not detected |
| Atlantic cod (<i>Gadus morhua</i>) | Not detected |
| Donkey (<i>Equus asinus</i>) | Not detected |
| Soya (<i>Glycine max</i>) | Not detected |
| Wheat (<i>Triticum aestivum</i>) | Not detected |

AA.2 Detection limit

The detection limit has been calculated upon standard samples consisting of mixtures of raw Ostrich meat and other species. [Imegen® Ostrich ID Kit](#) can detect blends containing a percentage below 0.01% (w/w) of Ostrich meat. The limit of detection in processed samples varies depending on the composition and food processing.

To ensure the representativeness of the results, we recommend the use of a DNA extraction method that allows you to process a large amount of sample (10-20 g). If you do not have a procedure with these features, we recommend the use of [Food Extraction Kit](#) (Part No: IMG-262).

AA.3 Quality certifications

- ✓ Health in Code, S.L. is certified against the standard UNE-EN ISO 9001:2015 "Quality management systems" for the design, development, manufacture, and commercialization of kits for genetic analysis.
- ✓ Health in Code, S.L. is certified against the standard UNE-EN ISO 14001:2015 "Environmental Management Systems" for the design, development, manufacture, and commercialization of kits for genetic analysis.

Appendix
B

Safety warnings and precautions



Strictly follow the instructions of this manual, especially regarding the handling and storage conditions.



Do not pipette by mouth.



Do not smoke, drink, or eat in areas where specimens or kit reagents are being handled.



You must properly protect any skin condition, as well as cuts, abrasions and other skin lesions.



Send down the drain only those materials found on the safe list. Compounds not listed are not suitable for drain disposal. Use waste containers according to the local legislation and manage their treatment through an authorized waste manager.



In case of an accidental release of any of the reagents, avoid contact with skin, eyes and mucous membranes and clean with abundant water.



The materials safety data sheets of all hazardous components contained in this kit are available on request to Health in Code, S.L.



This product could require the handling of samples and materials of human and animal origin. You should consider all human and animal source materials as potentially infectious and handled in accordance with OSHA Biosafety Level 2 of bloodborne pathogens or must use other relevant biosafety practices for materials containing or suspect that they may contain infectious agents.



Reagents included in this kit are non-toxic, neither explosive, infectious, radioactive, magnetic, corrosive nor environmental pollutants.



This kit has been validated with specific equipment under certain conditions, which could be different in other laboratories. It is recommended that each laboratory performs an internal validation when the kit is used for the first time.



The manufacturer is not responsible for the malfunction of the assay when one or more reagents included in the kit are replaced by other reagents not supplied by Health in Code, S.L.



The manufacturer does not guarantee the reproducibility of the assay when the user employs reagents not validated by Health in Code, S.L., considering them equivalent to those provided in the kit.



Documentation and support

AC.1 Food safety support

Please, visit our website for the latest services, orders and support information:


 imegenagro.es

Health in Code certificates of analysis and other product documentation:

 portal.imegen.es/en/certificate-of-analysis/

AC.2 Customer and technical support

For any questions about the applications of this product or this protocol, please contact our Technical Department:

 **+34 963 212 340**

 **tech.support@healthincode.com**

NOTE: For SDSs for reagents and chemicals from other manufacturers, please contact the appropriate manufacturer.
