



Instituto de Medicina Genómica SL

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All products sold by the Imegen are subjected to rigorous quality control. The **imegenTM Vegan ID kit** has passed all internal validation tests, ensuring the reliability and reproducibility of each assay.

For any questions about the applications of this product or its protocols, please contact our Technical Department:

Phone number: 963 212 340 **e-Mail**: info@imegen.es

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1. Product information

Kit description

Identification of meat species presence in food samples is an essential step in order to improve the traceability and the control within the food supply chain, as well as a necessary quality control for handling and cleaning processes of production lines.

Imegen-Vegan ID Kit allows determining the presence of animal DNA in any food by real time PCR.

Animal DNA detection is done by real time PCR using two TaqMan®-MGB probes. One of them, labelled with FAMTM dye, specifically detects one mitochondrial DNA sequence common in all animal species. The second probe is labelled with VIC® and detects an Internal Positive Control, which is used to rule out inhibitors in the sample and check the correct functioning of the assay.

The detection limit has been calculated upon standard samples consisting of mixtures of animal DNA and vegetal DNA. Imegen-Vegan ID Kit can detect blends containing a percentage below 0.01% of Animal DNA. The limit of detection in processed samples varies depending on the composition and food processing.

To ensure the representativeness of the results, we recommend the use of a DNA extraction method that allows you to process a large amount of sample (10-20 g). If you do not have a procedure with these features, we recommend the use of **Food Extraction Kit** (Part No: IMG-262).

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Content and storage of the kit

The kit contains needed reagents to perform the real time PCR reactions:

- Amplification primers for the PCR system.
- The system contains two probes labelled with FAM[™] and VIC[®] fluorophores, to detect the animal DNA and the Internal Positive Control (IPC).
- Vegan Positive Control (0,1% of Bos Taurus DNA)
- 2X General Master Mix.

The kit contents the necessary reagents to perform 48 reactions:

Reagents	Color	Amount	Storage
Vegan Master-Mix	Black pad	360 µl	-20°C
Vegan positive control	Orange cap	120 µl	-20°C
General Master-Mix	White pad	600 µl	4°C

Table 1. Kit components and storage temperature of imegen-Vegan ID Kit

This kit has been proved and is compatible with the following real time PCR platforms: 7500 FAST and StepOne Real-Time PCR System (Thermo Scientific).

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Equipment and material required

In the following table the equipment requirements for using **imegen-Vegan ID Kit** are shown:

Equipments			
1	Real-time PCR Thermal Cycler with channels for detection of FAM™ (520 nm) and VIC® (550 nm)		
2	Micropipettes (10 μL, 20 μL and 200 μL)		
3	Table top centrifuge with adaptors for 96 well PCR plates and/or 0,2 ml tubes		
4	Vortex		

Materials			
1	Optical 96-well reaction plates or 0.2 ml optical tubes		
2	Optical adhesive film for 96 well plates or optical caps for 0.2 ml tubes		
3	Disposable micropipette filter tips (10 $\mu L,~20~\mu L$ and 200 $\mu L)$		
4	1.5 mL sterile tubes		
5	Powder-free latex gloves		

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2. Real Time PCR

PCR reactions preparation

A PCR master mix should be prepared in order to perform the analysis with imegen-Vegan ID kit:

The PCR Master Mix contain:

- Vegan Master Mix
- General Master-Mix (2X)

The recommended protocol for preparation of amplification reactions is showed below:

- 1. Thaw the Vegan Master-Mix, Vegan Positive Control and samples DNA.
- 2. Vortex each reagent and keep cold.
- 3. Add into a 1.5 mL tube the following reagents (table 2). To estimate the amount of necessary reagents, we recommend make calculations taking into account the number of samples to be simultaneously analysed, and then considering one more reaction, or increase a 10% the volume of each reagent.

Reagents	Amount per reaction
Vegan Master-Mix	7.5 µL
General Master Mix	12.5 μL

Table 2. Reagents amount per reaction in 7500 FAST or StepOne

4. Vortex the PCR Master Mix tube and dispense 20 μL into corresponding wells.

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- 5. Add 5 µl of each DNA sample at 10 ng/µl, 5 µl of Vegan Positive Control and 5 µl of the Negative Controls* into the appropriate wells.
 - We recommend making each sample analysis in duplicate.
- 6. Cover the well plate with optical film or the tubes with optical cover and spin in the centrifuge.
 - * We strongly recommend using an extraction negative control for each run of extractions carried out. This control consists in one tube to which no sample is added and which is summited to the same extraction process as the other samples. Likewise, we recommended using a PCR negative control for each PCR run; this tube contains no DNA but all PCR reagents.

PCR amplification program

This kit is compatible with the Real-time PCR platforms 7500 FAST, StepOne Real-Time PCR System (Thermo Scientific), and any other Real-time PCR platforms equipped with FAM and VIC channels.

Probes	Receptor	Quencher
Animal-P	FAM TM	MGB
IPC-P	VIC®	MGB

Table 4. Probes information

The following instructions should be taken into account in order to setup the amplification program:

Reaction volume: 25 μL

Targets: FAM and VIC

 In case the quencher has to be defined, select MGB for all probes. If the real time PCR system does not take into account the quenchers, select only the

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receptors (FAM and VIC).

- If the real time PCR system is a 7500 Fast or a StepOne Real-Time PCR system [Thermo Scientific] select Quantitation- Standard curve as a type of experiment and include ROX[™] as a reference.
- Ramp rate: standard
- Optimal program:

Fields	Step 1 Enzyme activation		ep 2 PCR
Cycle		36 cycles	
Number	1 initial cycle	Denaturation	Primers binding/Extention
Temperature	95°C	95°C	60°C
Time	10 minutes	15 seconds	1 minute*

Table 5. Optimal PCR program

Note: This program has been validated on a StepOne Real-Time PCR System from Applied Biosystems. If you use another brand or model of thermal cycler, you may need the amplification program to be adjusted. Please contact our service department for advice.

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^{*}Fluorescence detection



3. Analysis of results

Before analysing the samples results, you should stablish the threshold in 0.4 and, also, you should check that the results obtained in the controls, is as expected:

- **Positive Control**: The result must always be positive in all amplification reactions, both in the FAM^{TM} channel as VIC^{\otimes} .
- Negative controls: Amplification must be only detected in the VIC[®] channel.
 In this channel an internal positive control [IPC] is detected, which determines the absence of inhibition in the sample.

IPC

It must be checked that the IPC (VIC[®]) is positive in all samples, with a Ct similar to the Positive Control. A negative result in the IPC indicates the presence of inhibitors in the sample. It should be noted that IPC result may be negative in samples where a lot of animal DNA (FAMTM) is detected, because the PCR reagents are exhausted before amplification of the IPC begins.

Animal DNA

Amplification in the FAM^TM channel indicates presence of animal DNA in the sample.

It is necessary to check if sample Ct is less than the Ct_{cut-off} in order to determine if one reaction of amplification is positive. Any reaction of amplification with Ct upper than Ct_{cut-off} may be considered as negative. The Ct_{cut-off} is equal than the positive control Ct (0.1%) plus 3.32.

$$Ct_{cut-off} = 3.32 + Ct_{Positive\ Control}$$

Note: Any sample with a Ct equal than Ct_{cut-off} contains approximately 0.01% of animal DNA.

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In samples where no amplification in the FAM^{TM} channel is seen, we can conclude that no animal DNA are detected or that their amount in the sample is below than the detection limit.

The following table shows graphically the results that may be obtained from one sample analysis, as well as the interpretation that should be done from the obtained result:

Vegan Master Mix			
Animal	IPC	Interpretation	
-	+	No animal DNA are detected	
+	+	Animal are detected	
-	-	PCR inhibitors presence in the sample*	
+	-	Sample with big amount of Animal DNA	

Table 5. Results interpretation

The following table shows graphically the results that may be obtained from the analysis of different assay controls, as well as the interpretation that should be done from the obtained result:



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^{*} If presence of inhibitors in the sample is detected, we recommend checking whether there has been an excess of DNA in the reaction (the recommended maximum is 250 ng). If the amount of DNA is right, we recommend repeating DNA extraction. If the problem persists, please contact our technical department.



Controls	Vegan Master Mix			
Controls	Animal	IPC	Interpretation	
Positive	+	+	Expected result	
control	-	-	PCR Amplification Failure ¹	
Extraction	-	+	Expected result	
negative control	+	+	Contamination in the DNA extraction procedure ²	
PCR	-	+	Expected result	
negative control	+	+	PCR contamination with animal DNA ³	

Table 6. Interpretation of possible results

Recommendations:

- PCR Amplification Failure: Check amplification program and configuration of fluorescence capture. Amplification failure may be due to a setup technical problem.
- ² Contamination in the DNA extraction procedure: Contamination may be due to some error made in the process of sample handling, reagents contamination, or environmental contamination. Check DNA extraction protocol, wipe the laboratory where DNA extraction process was performed and take care to avoid any contamination during sample homogenization. If necessary, use new aliquots of the reagents used in DNA extraction.
- ³ PCR contaminations with animal DNA: Contamination of PCR reactions may be due to an error made in the process of sample handling, contamination of the reagents or environmental contamination. Thoroughly clean the laboratory where the PCR process was performed, as well as equipment. If necessary, use new aliquots of the reagents used in the PCR. Prepare the PCR reaction containing the Positive Control last to avoid cross contamination.

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